

## QuinDye™ Terminator

Catalog No: CPR00001,CPR00002,CPR00003,CPR00004

**For Research Use Only. Not for use in diagnostic procedures.**

### Product Description

QuinDye™ Terminator Kit provides premixed reagents for Sanger sequencing reactions. The kit includes Dye Terminator and a 5x buffer, formulated to deliver long, high-quality reads with uniform peak heights and to efficiently read through complex sequencing structures.

### Components/Kit Information

Catalog No	CPR00001	CPR00002	CPR00003	CPR00004
Reactions	24 Reactions	100 Reactions	1000 Reactions	5000 Reactions
QuinDye Sequencing Mix	192 µL*1	800 µL*1	800 µL*10	20 mL*2
pGEM control DNA	10 µL*1	10 µL*1	250 µL*1	250 µL*2
M13 (-21) control primer	10 µL*1	10 µL*1	200 µL*1	200 µL*2
QuinDye 5x Sequencing Buffer	1 mL*1	1 mL*2	12 mL*1	28 mL*2

### Storage Condition

Store QuinDye Sequencing Mix, pGEM control DNA, and M13 (-21) control primer at –30°C to –10°C in a frost-free freezer. Protect the sequence mix from light. Store QuinDye 5x Sequencing Buffer at 4°C in a frost-free freezer

### REAGENTS to be supplied by the User

- Reagent Alcohol, EtOH
- Nuclease free H<sub>2</sub>O

### Compatible Sequencing Instruments

- 3730 DNA Analyzer
- 3500 Genetic Analyzer
- 3130 Genetic Analyzer

### Procedure

## 1. Prepare DNA Template

DNA template	Recommended Quantities
PCR product	10-20ng per kb
Single-stranded DNA	25–50 ng
Double-stranded DNA	150–300 ng
Cosmid, BAC	0.5–1.0 µg
Bacterial genomic DNA	2–3 µg

Notice, the sequencing templates should be purified before use in sequencing reactions.

## 2. Perform cycle sequencing

### 2.1 Set Up the Sequencing Reactions

**2.1.1** Completely thaw the components of the QuinDye™ Terminator Kit and your primers, then store them on ice.

**2.1.2** Vortex the tubes for 2 to 3 seconds, then centrifuge 2 to 3 seconds to collect contents at the bottom of the tubes.

**2.1.3** Add components as indicated. This setup is for standard 20 µL reactions, but the volume can be reduced to 10 µL in 384-well plate. In this case, maintain the same primer concentration and volume as used in the 20 µL reactions.

**2.1.4** Seal the plate and vortex for 2 to 3 seconds, then briefly centrifuge to collect contents at the bottom of the wells (5 to 10 seconds) at 1,000 × *g*.

Component	Quantity per reaction	Example Forward	Example Reverse
QuinDye Sequencing Mix	8 µL	8 µL	8 µL
Forward primer (3 µM)	1 µL	1 µL	-
Reverse primer (3 µM)		-	1 µL
Deionized water (RNase/DNase-free)	Varies based on template and primer volume	9 µL	9 µL
Template	See above template quantity*	2 µL	2 µL
<b>Total volume</b>	<b>20 µL</b>	<b>20 µL</b>	<b>20 µL</b>

### 2.2 Run the sequencing reactions

**2.2.1** Place the tubes or plate(s) in a PCR machine and set the volume.

**2.2.2** Perform cycle sequencing:

Parameter	Stage/step				
	Incubate	Cycling (25 cycles)			Hold
		Denature	Anneal	Extend	
Ramp rate	—	1°C/second			
Temperature	96°C	96°C	50°C	60°C	4°C
Time (mm:ss)	01:00	00:10	00:05	04:00*	Until ready to purify

\* Shorter extension times can be used for short templates.

### Using QuinDye™ Terminator 5X Sequencing Buffer to dilute sequencing reactions

Some cycle sequence reactions may be optimized using diluted QuinDye™ Sequencing Mix. The QuinDye 5x Sequencing Buffer can be used to dilute the QuinDye Sequencing Mix.

Note: If you use the QuinDye 5X Sequencing Buffer without optimization, the quality of the sequence may deteriorate. We can not guarantee the performance of QuinDye™ chemistry when it is diluted.

An example of a 0.5x diluted sequencing reaction is shown below:

Component	Quantity per reaction	Example Forward	Example Reverse
QuinDye Sequencing Mix	4 µL	4 µL	4 µL
QuinDye 5x Sequencing Buffer	2 µL	2 µL	2 µL
Forward primer (3 µM)	1 µL	1 µL	-
Reverse primer (3 µM)		-	1 µL
Deionized water (RNase/DNase-free)	Varies based on template and primer volume	11 µL	11 µL
Template	See above Template quantity*	2 µL	2 µL
<b>Total volume</b>	<b>20 µL</b>	<b>20 µL</b>	<b>20 µL</b>

Concentration of template may affect volume, if template volume differs please adjust the volume of water in the reaction mix.

**IMPORTANT! Protect dye terminators from light. Cover the reaction mix and sequencing plates with aluminum foil before use.**

For technical support, please visit [Quintara Bioscience](http://Quintara Bioscience) or email technical support team [sales.us@quintarabio.com](mailto:sales.us@quintarabio.com)

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